

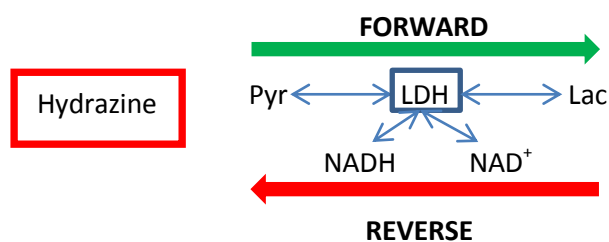
Hons practical MC 4

GROUPS:

- 1A**
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 - Greta De Waal
- 2B**
- Melinda Badenhorst
 - Yasmin Masoudi
- 3C**
- Nicole Green
 - Tammy Speelman
- 4D**
- Stefan Botha
 - Will de Villiers
- 5A**
- Kay Leukes
 - SinJon Heitmann
- 6B**
- Ané Louw
 - Lauren Myburg
 - Anna-Mart Burger

ABOUT THE PRAC:

(1) Characterise the enzyme:



(2) The experiments:

GROUP A

- Reverse direction
- Varying [NAD⁺]
- Same [Lac] + [LDH] + [Hydrazine]

GROUP B

- Reverse direction
- Varying [Lac]
- Same [NAD⁺] + [LDH] + [Hydrazine]

GROUP C

- Forward direction
- Varying [NADH]
- Same [Pyr] + [LDH]

GROUP D

- Forward direction
- Varying [Pyr]
- Same [NADH] + [LDH]

GROUP A

- Product Inhibition: NADH
- Varying [NADH]
- Same [Lac] +[NAD⁺] + [LDH] + [Hydrazine]

GROUP B

- Without Hydrazine
- Varying [Lac]
- Same [NAD⁺] +[LDH]

GROUP C

- Product Inhibition: NAD⁺
- Varying [NAD⁺]
- Same [Pyr] +[NADH]+[LDH]

GROUP D

- Product Inhibition: Lac
- Varying [Lac]
- Same [NADH] + [Pyr]+[LDH]

(3) An enzyme assay:

- a- Pipette metabolites into the cuvettes, the reaction is initiated by the addition of the enzyme.
- b- Once the enzyme is added measure the absorbance readings for the reaction over a period of 3mins.
- c- The types of data we expect to see- Forward reaction: decreasing Abs; Reverse reaction: increasing Abs

(4) What needs to be done:

Create a pipetting schema so that we investigate 10 different concentrations around the Km values. Take the following into consideration:

a- Given :

Km	Stock concentrations
Km(NADH) = 0.05 mM	[NADH] = 1.5 mM
Km(NAD ⁺) = 0.4 mM	[NAD] = 5 mM
Km(Pyr) = 0.5 mM	[Pyr] = 10 mM
Km(Lac) = 10 mM	[Lac] = 1 M

HYDRAZINE is given at a 10X dilution of bottle concentration

b- Volumes:

- The total cuvette volume is 1 ml and of that you use 10 µl of the enzyme (LDH) and if you are doing **reverse direction experiments (only)** 35 µl is the Hydrazine.
- The other volumes are of the substrate and co-factor which you calculated in the pipetting schema.
- Lastly, the remaining volume to make up this 1 ml volume is done by adding buffer to the cocktail mixture.

c- How to determine the 10 concentrations:

- The range you want to investigate is between: [Km/10; Km] and (Km; 10Km] and the fixed substrates are usually at saturating condition ie. 10xKm. Use Excel to create an incremental pipetting schema.

(5) The Program:

18/04 Tuesday:	19/04 Wednesday:	20/04 Thursday:
Morning: Practical theory and pipetting schema	Morning: Group 4-6 (first set of experiments)	Morning: 09H30 Research lecture
Afternoon: Group 1-3 (first set of experiments)	Afternoon: Group 1-3 (second set of experiments)	Mid morning: Group 4-6 (second set of experiments)

21/04 Friday: Morning: Data pooling/ initial data analysis Afternoon: redo experiments?	24/04 Monday: Morning: Data analysis/ Redo experiments?	25/04 Tuesday: Data analysis day
26/04 Wednesday: Data analysis day	27/04 Thursday: FREEDOM DAY (PUBLIC HOLIDAY)	